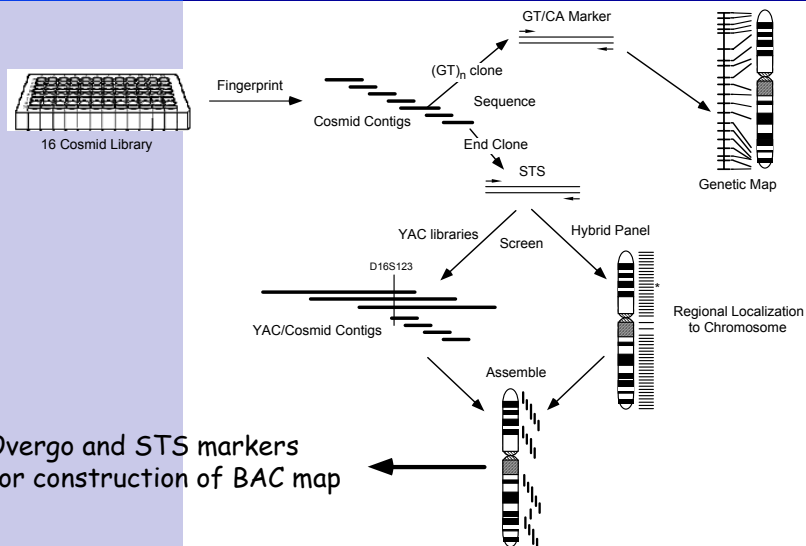
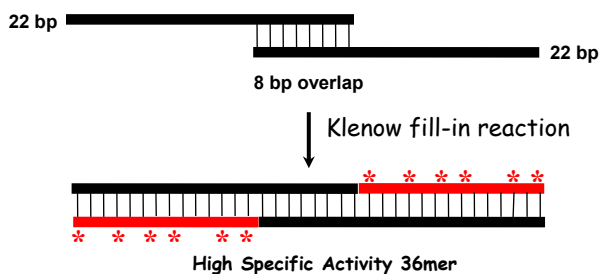


## YAC/STS/Cosmid Map of Chromosome 16



Integrated Physical Map  
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## Overgo Probe

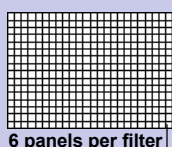
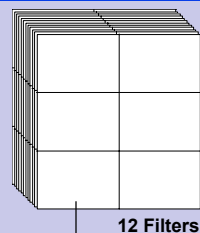


### Features

1. Easily designed to be unique
2. Simple to handle
3. Suitable for complex pooled hybridizations
4. High specific activity labeling

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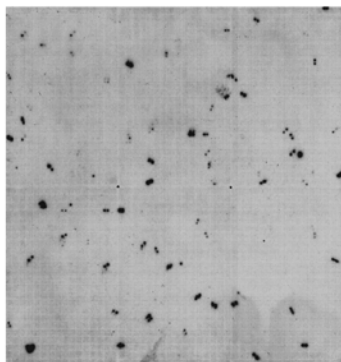
## Top pool hybridization



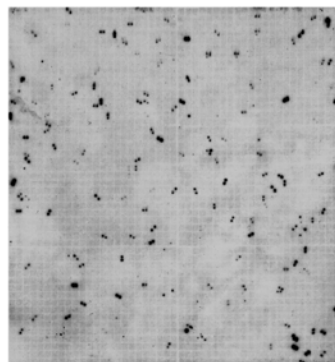
24 x 16  
blocks of  
4x4  
colonies  
221,184 BAC clones



Pod name	1	2-9	3-4	5-6	7-8-10	Total
Number of STSs	96	192	192	192	236	908



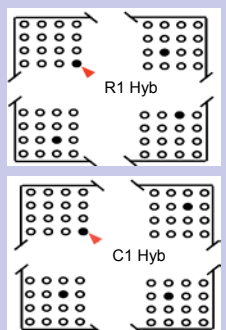
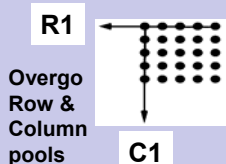
96 overgos



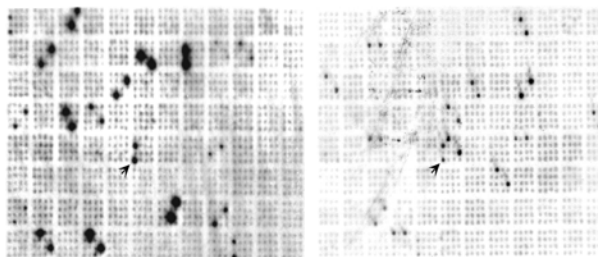
192 overgos

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## Two dimensional hybridization



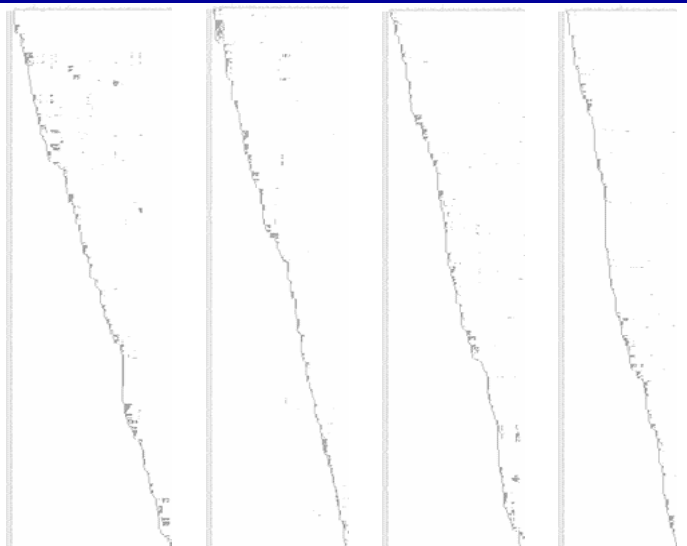
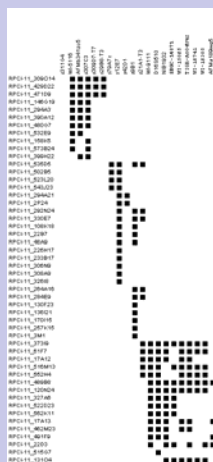
Group	YAC	BRPT	WI	SHGC	C16	Total
Number of STSs	402	128	166	64	148	908
Column pools	24	24	24	24	24	120
Row pools	17	6	8	3	7	41



Left: Pool Column 17 of WIC group; filter number 5.  
Right: Pool Row 6 of WIC group; filter number 5. The overgo at the intersection of the two pools is WI-14879.

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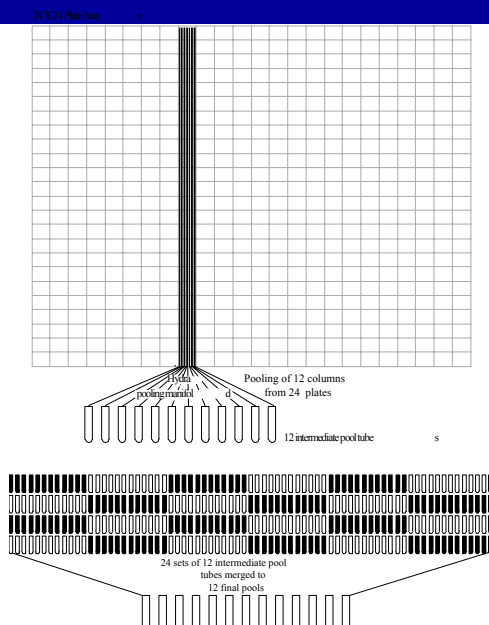
## Probe-content Maps for 16q

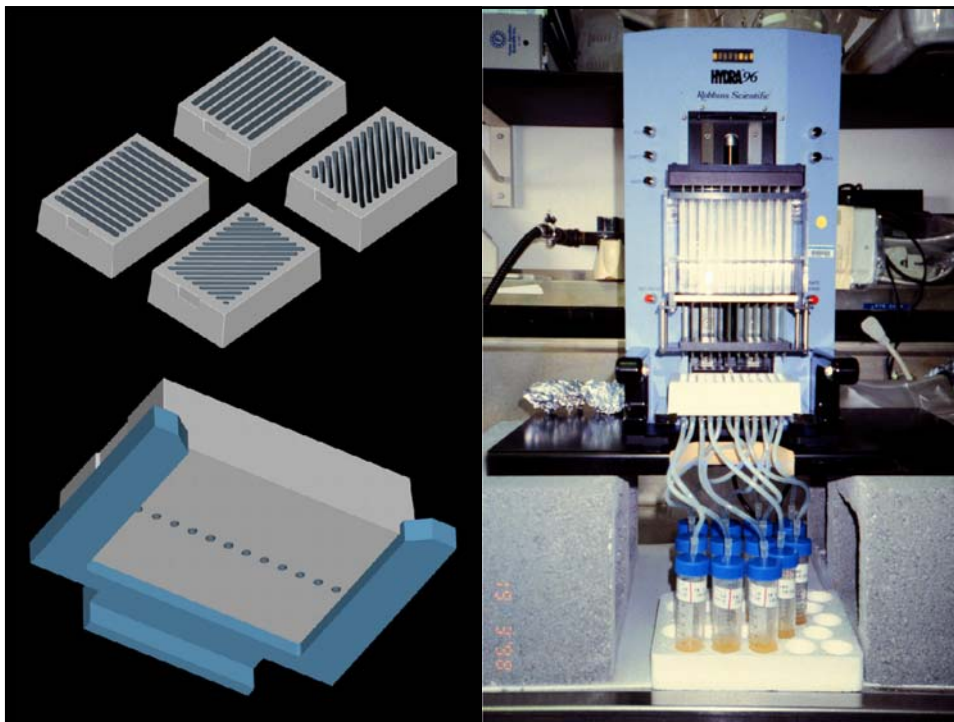


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## BAC Library Pools for PCR Screening

- 8 dimensional single tier design
- pools consist of 2 sets each of plate pools, row pools, column pools and diagonal pools
- pools are constructed using Robbins Hydra and appropriate manifolds





## BAC end walking

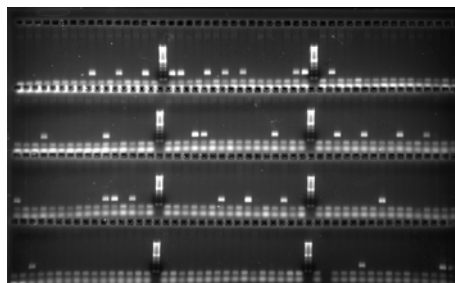
Clones adjacent to gaps



BAC end sequences



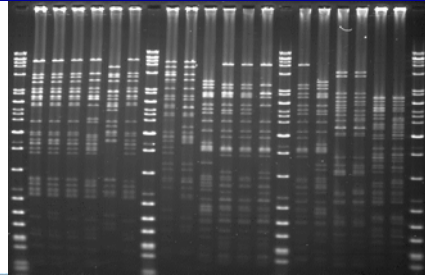
Design PCR primer and screening



Agarose gel image for screening a single STS against the single 376 pools. Marker lanes are included at 8 positions on the gel.

## Restriction map assembly

All BACs are fingerprinted with EcoR I.



Restriction maps  
Are constructed  
Using GRAM



Minimal tiling  
set clones are  
selected for  
sequencing.

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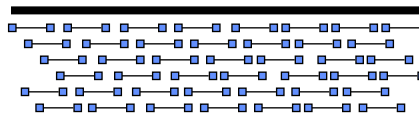
## Double End Plasmid Sequencing

5X double end plasmid  
sequencing of 3 kb  
subclones

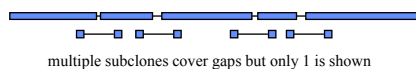
98% sequence coverage  
100% subclone coverage

100% sequence coverage

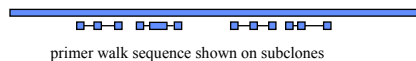
Clone



Assembled  
Sequence



Primer  
Walking

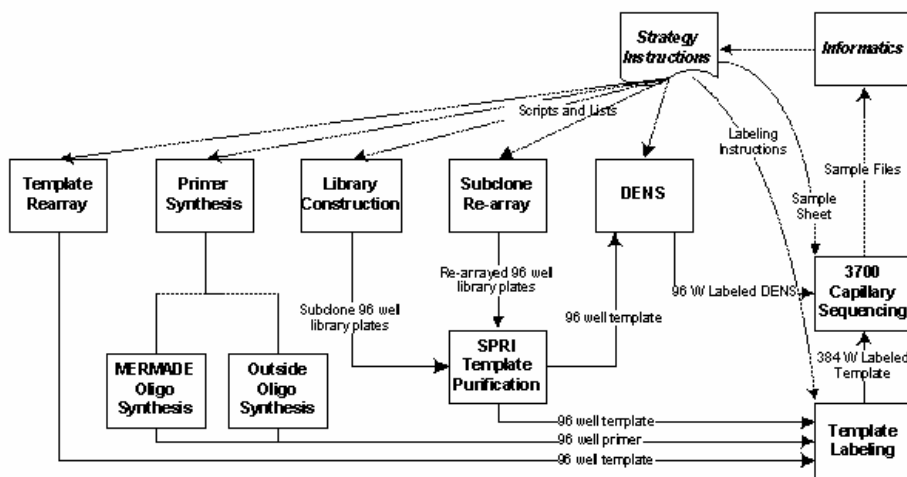


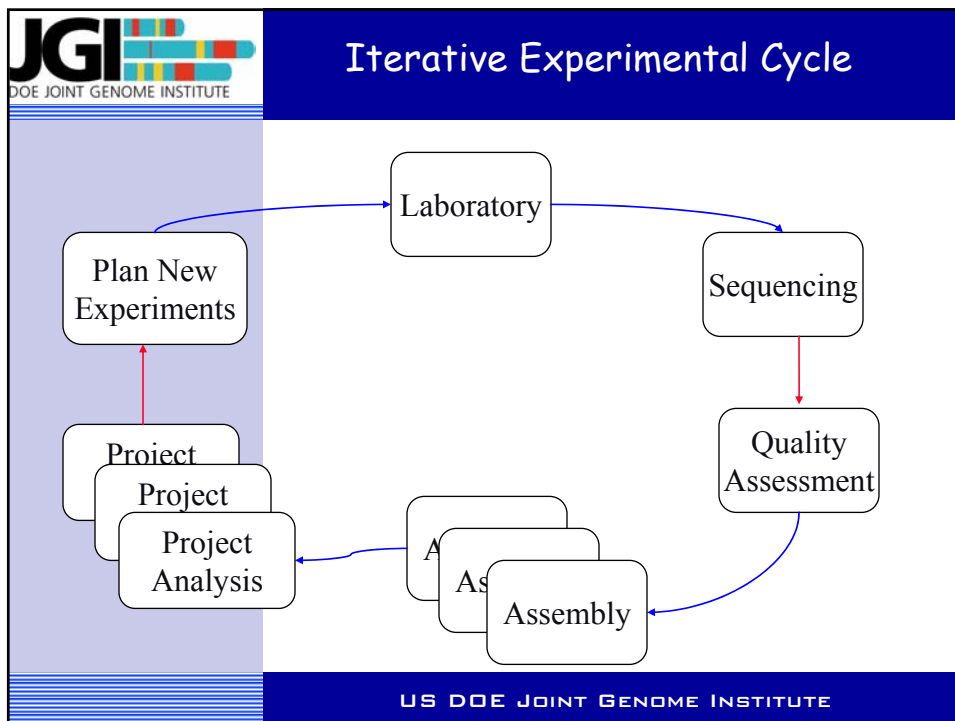
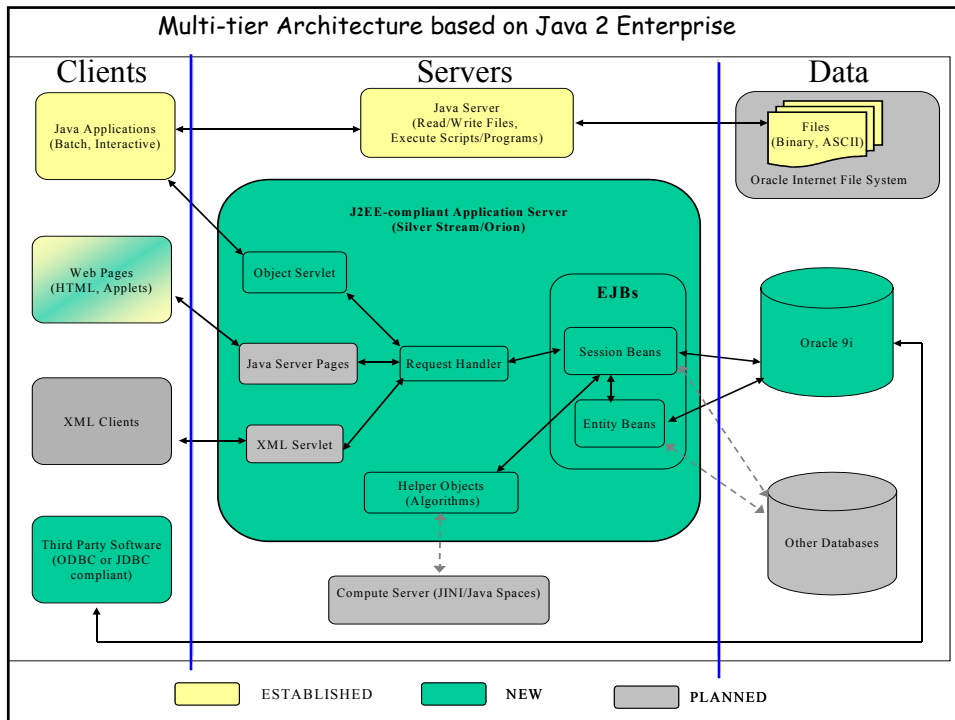
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## Whole Chromosome Finishing

- All BAC projects for the chromosome are considered simultaneously.
- Sequence data is merged from overlapping projects.
- A finishing target in an overlapping region is considered once.
- An Oracle database maintains records of all finishing reactions for the whole chromosome and tracks these through the finishing process.

## Informatic Automation Drives all steps of Finishing





## Generation of Finishing Reactions

- Primarily batch-oriented with GUI interface as necessary
- Create reactions: dp, dgtp, custom primers, dens, enhancer, shatter libraries
- Group reactions into plates by priority, source plate, primer temperature, etc
- Generate appropriate robot files

## Complete Project Management

The screenshot displays three windows from the JGI Complete Project Management software interface.

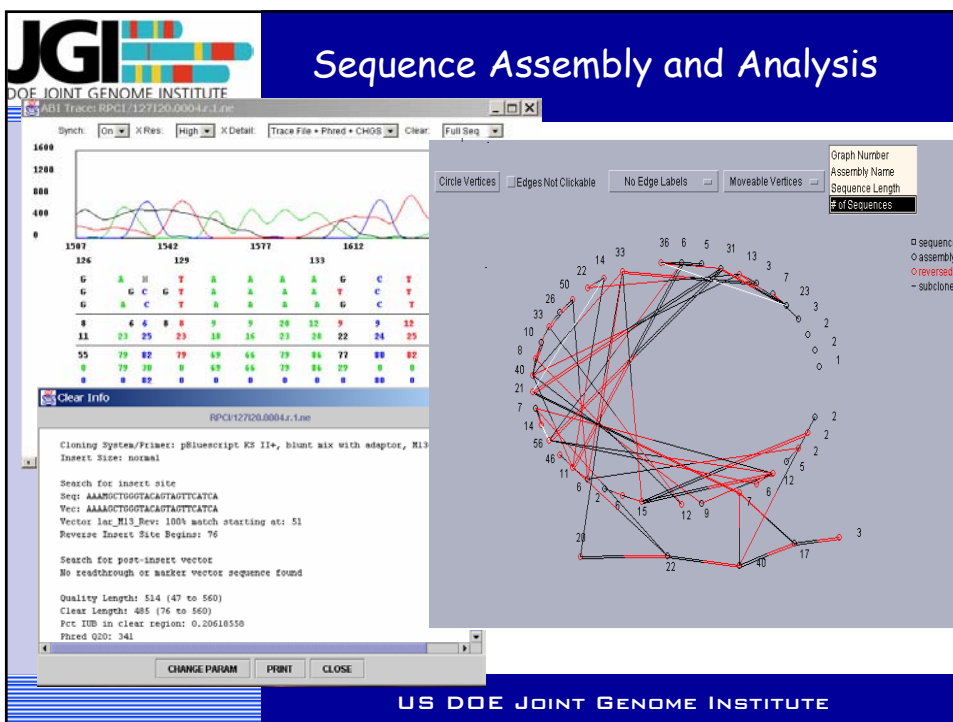
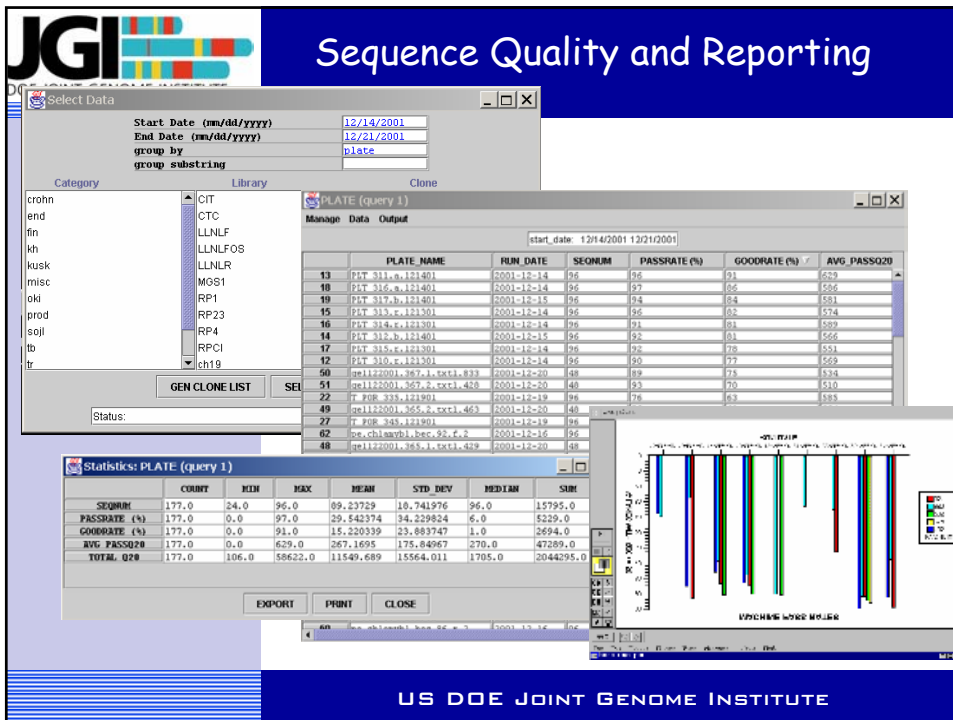
**Update Reactions Window:** A table showing reaction plate names and their status.

REACTION PLATE NAME	STATUS
DENS_318	In Progress
DENS_309	In Progress
T POR_333	In Progress
DENS_320	In Progress
DENS_321	In Progress
DENS_322	In Progress
DENS_323	In Progress
T POR_336	In Progress
DENS_325	Complete
T POR_334	In Progress
DENS_329	In Progress
T POR_335	In Progress
DENS_336	In Progress

**Select Reaction Plates Window:** A window for selecting reaction plates for four different plates (A, B, C, D). Each plate has a list of reaction names and a suffix field.

**Generate Sample Sheet Window:** A window for generating a sample sheet. It includes a table of sequence names and a dropdown menu for sample names.

SEQUENCE NAME	general properties	PROJECT
A-B1	2270F3.2047.p.i.de	
A-B1	2270F3.2048.p.i.de	
A-C1	2270F3.2049.p.i.de	
A-D1	2270F3.2050.p.i.de	
A-E1	2270F3.2051.p.i.de	
A-F1	2270F3.2052.p.i.de	
A-G1	2270F3.2053.p.i.de	
A-H1	2270F3.2054.p.i.de	
A-I1	2270F3.2055.p.i.de	
A-J1	2270F3.2056.p.i.de	
A-K1	2270F3.2057.p.i.de	
A-L1	2270F3.2058.p.i.de	
A-M1	2270F3.2059.p.i.de	
A-N1	2270F3.2060.p.i.de	
A-O1	2270F3.2061.p.i.de	
A-P1	2270F3.2062.p.i.de	
A-Q1	2270F3.2063.p.i.de	
A-R1	2270F3.2064.p.i.de	
A-S1	2270F3.2065.p.i.de	
A-T1	2270F3.2066.p.i.de	
A-U1	2270F3.2067.p.i.de	
A-V1	2270F3.2068.p.i.de	
A-W1	2270F3.2069.p.i.de	
A-X1	2270F3.2070.p.i.de	
A-Y1	2270F3.2071.p.i.de	
A-Z1	2270F3.2072.p.i.de	
A-1	2270F3.2073.p.i.de	
A-2	11686.2770.p.i.de	
A-3	11686.2771.p.i.de	
A-4	11686.2772.p.i.de	
A-5	11686.2773.p.i.de	



## Finishing Strategy

- Finishing Reactions**
- Big Dye reactions sequenced by ABI 3700
  - dGTP Big Dye terminator reactions
  - Invitrogen SequencerRx reaction enhancer or other enhancer
  - Shatter libraries
  - Alternate reaction chemistries
  - Li-Cor infrared Global IR2 platform

- **Combine Long Reads, Redos & Primer Walks**
- **Targets & Reactions**
  - **Chemistry/Strategy Based on Desired Read Length**
  - **Geometry First for Strands**
  - **Quality Second**
  - **Priority and Alternate Reactions**
  - **Database Tracking for Efficient Rearray**

## Primer Walks

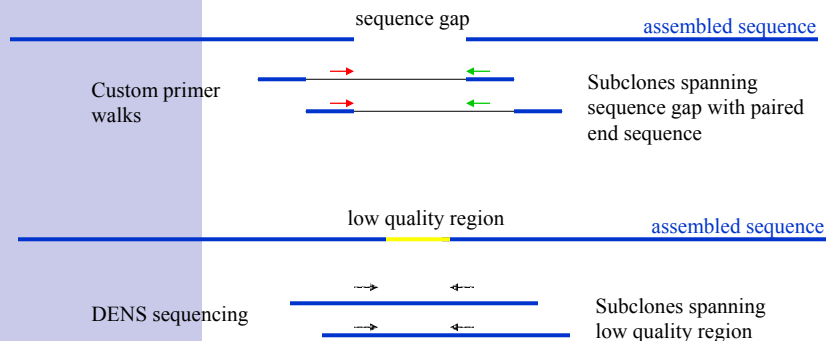
### Custom Primers

- (Mermade) to Close Gaps
  - Automation for 2 Subclones/Primer
    - Q BOT Correlated Rearray
    - Half-plate Hydra Liquid Primer Handling
  - Database Generates and Tracks Primer Rxn's

### DENS

- (Octomer library) for Sequence Quality Improvement
  - Database Tracks Octomer Usage and Stat's

## Custom Primer Walks and DENS



## DENS (Differential Extension with Nucleotide Subsets)

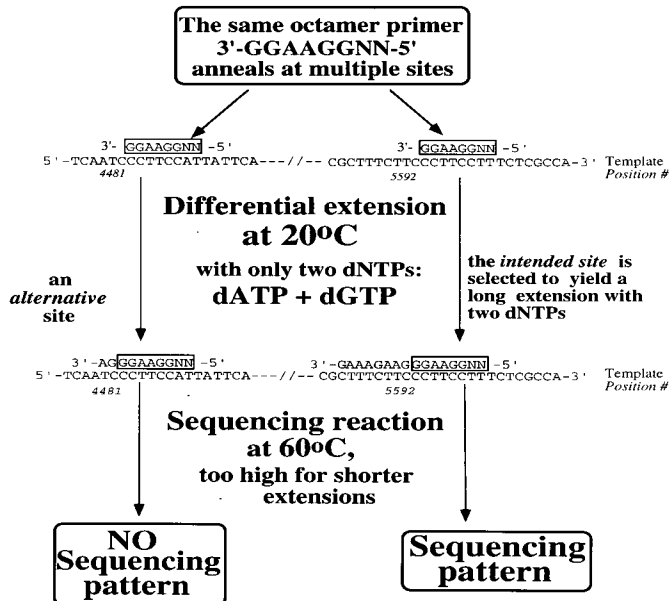
Primer walking without  
custom primer synthesis

**NNAGGCTA**

Raja et al., NUCLEIC ACIDS RESEARCH, 25:800-805 (1997) Zevin-Sonkin et al., DNA SEQUENCE, 10:245 (1999)

## DENS mechanism

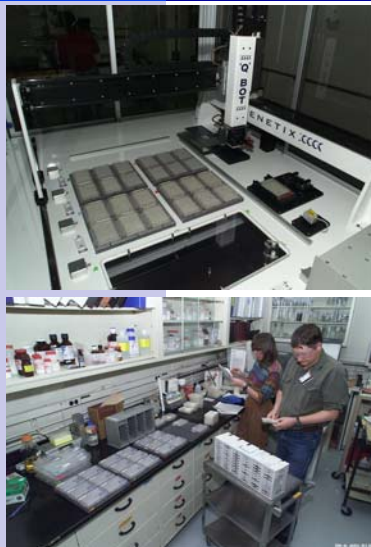
DENS works by converting a short primer (selected from a pre-synthesized library of 1440 double-degenerate octamers) into a long one on the template at the *intended site only*



## Features of Finishing Pipeline

- 96 well Q-Bot clone re-array
- 96 well SPRI template prep
- 96 well MerMade oligonucleotide synthesis
- 384 well sequencing reaction using 1/6X reaction and 96 channel liquid handling
- ABI 3700 96 capillary format and automatic loading

## Automation for Subclone Re-array



- Genetix Q-Bot
- Custom software to permit skips for controls, duplicating wells and flexible positioning.
- Database initiation and tracking
- 384 well compatible
- 60 re-arrayed destination plates per week

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## SPRI DNA Preps

- Up to 240 96 well plates per week
- On universal primer, terminator chemistry, template produces pass rates of ~90% and read lengths > 650 BP



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## Template Labeling



- Up to 80 384 well plates labeled per week
- Over 5,000 plates labeled per year
- Four 96 well template plates are merged to one 384 well plate
- Chemistries
  - Custom primer or universal, Big Dye terminator
  - Big Dye primer
  - Universal or custom primer, dGTP Big Dye terminator

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## MerMade Oligonucleotide Synthesis



- Up to 20 plates synthesized per week with two MerMades
- 96 well synthesis format
- Purity confirmed with MALDI TOF Mass Spec and gel electrophoresis
- Upgraded chemical ventilation system
- Developing protocol for Universal CPG to simplify synthesis plate

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## Sequencing Instrumentation



- Up to 200 ABI 3700 runs per week
- Five ABI 3700 capillary sequencers
- Five ABI 377 slab gel sequencers
- One Amersham MegaBACE 1000 capillary sequencer

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## Human Chromosome 16 January 29, 2002

### Chr16 size

Cytogenetic estimate  
98 Mb total  
89 Mb euchromatin  
Celera Scaffolds  
81 Mb  
TPF Unique restfrags  
77.6 Mb

#Cosmids, BACs,  
P1's, PACs and  
YAC

### Finished TPF Clones

Total	63.5 Mb	462 Clones <sup>#</sup>
Unique	~52 Mb	(~58%)

### In Finishing\*

(at LANL):	37.22 Mb	216 BACs
------------	----------	----------

### In Drafting at PGF\*

Depth (>6x):	4.77 Mb	30 BACs
In RCA (0x):	2.40 Mb	15 BACs
Glycerol (0x):	0.67 Mb	8 BACs & Cosmids

\*Mb are sums of total clone sizes (not unique)

Clone Gaps (type-3):	13
Estimated remaining BACs:	18

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## LANL Finishing Progress To Date

	Month	# Clones	Total	Unique
	1997	2	70,679	70,679
	1998	43	2,304,776	2,031,393
	1999	9	1,279,964	1,273,078
	2000	20	3,376,785	3,210,578
	Jan-Jul	9	1,589,347	1,269,293
	Aug'01	13	2,142,612	1,613,169
	Sep'01	23	4,260,548	3,431,062
	Oct'01	31	4,899,475	3,372,520
	Nov'01	24	3,865,978	2,894,949
	Dec'01	25	3,809,468	2,269,030
	Jan'02	65	10,857,259	~7,500,000
	<b>Total</b>	<b>264</b>	<b>38,456,891</b>	<b>~29,000,000</b>

Who Drafted  
What LANL  
Finished? **Draft** / Finish  
**Center** / Center

17.4 Mb **PGF**/LANL  
15.1 Mb **LANL**/LANL  
5.9 Mb **G5-PGF**/LANL

**77.7% Unique**  
(through Dec'01)

## Human Chromosome 16 TPF

### TPF Map Status

731 total clones  
32 clones w/o accessions (9 for PCR only)  
13 type-3 gaps  
No redundancies with other chromosomes

Clone type	Finished	Drafted	In draft	Total	% Finished
RP11 BAC	250	176	9	435	57.5%
Caltech BAC	115	57	17	189	60.8%
Cosmid	89	4	0	93	95.7%
PAC	5	3	0	8	62.5%
P1	4	0	0	4	100.0%
YAC	1	0	0	1	100.0%
PCR Fragment	0	0	1	1	0.0%
<b>Total</b>	<b>464</b>	<b>240</b>	<b>27</b>	<b>731</b>	<b>63.5%</b>

## Human Chromosome 16 TPF

The TPF (tiling path file) is the minimal tiling set of clones that provides complete coverage of the chromosome.

Redundancy estimated as follows:

89 Mb chr16  
-2.1 Mb gap  
= 86.9 Mb target  
108.08 Mb coverage/  
87.9 Mb target = 1.244

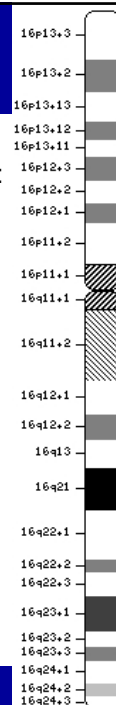
### TPF Statistics

<b>Clone distribution:</b>	<b>Unique Coverage:</b>
437 RPCI-11 BACs	32.0 Mb p-arm
189 CalTech BACs	45.6 Mb q-arm
93 Cosmids	6 p-arm gaps
7 PACs	7 q-arm gaps
4 P1's	
1 YAC	

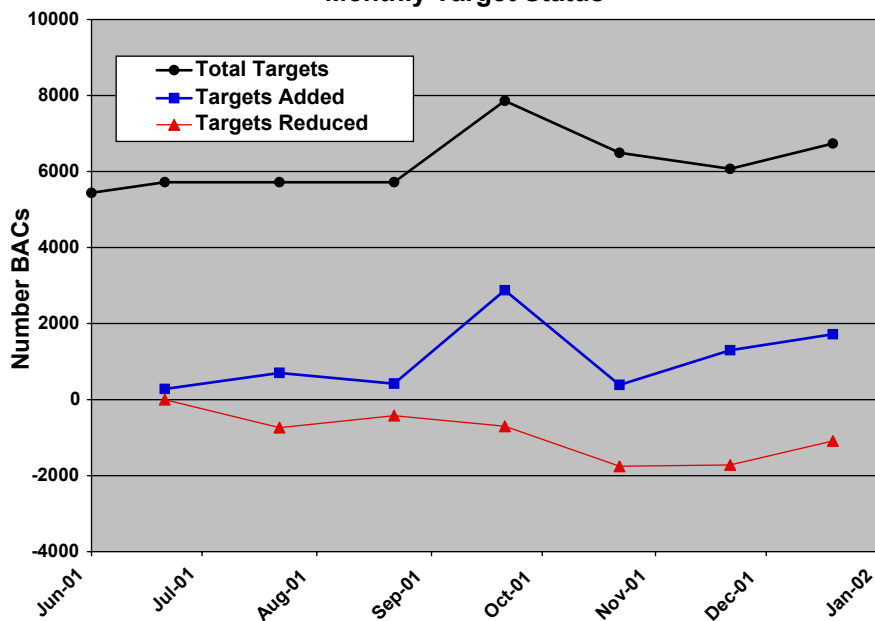
Total clone size	108,079,774 bp
Total gap size	2,100,000 bp
Estimated redundancy in TPF	1.244

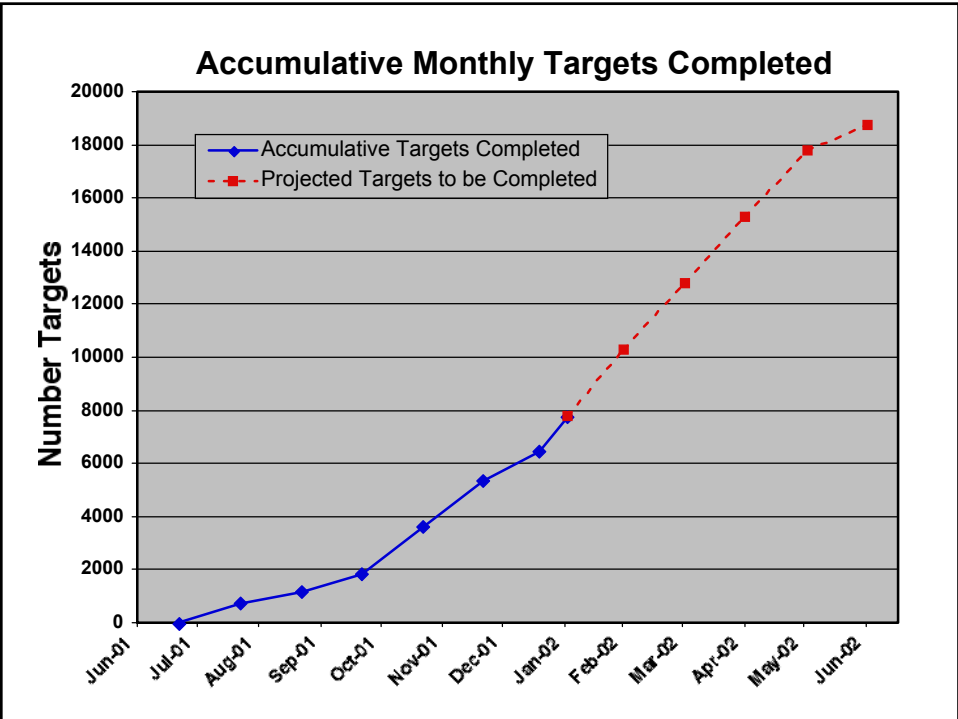
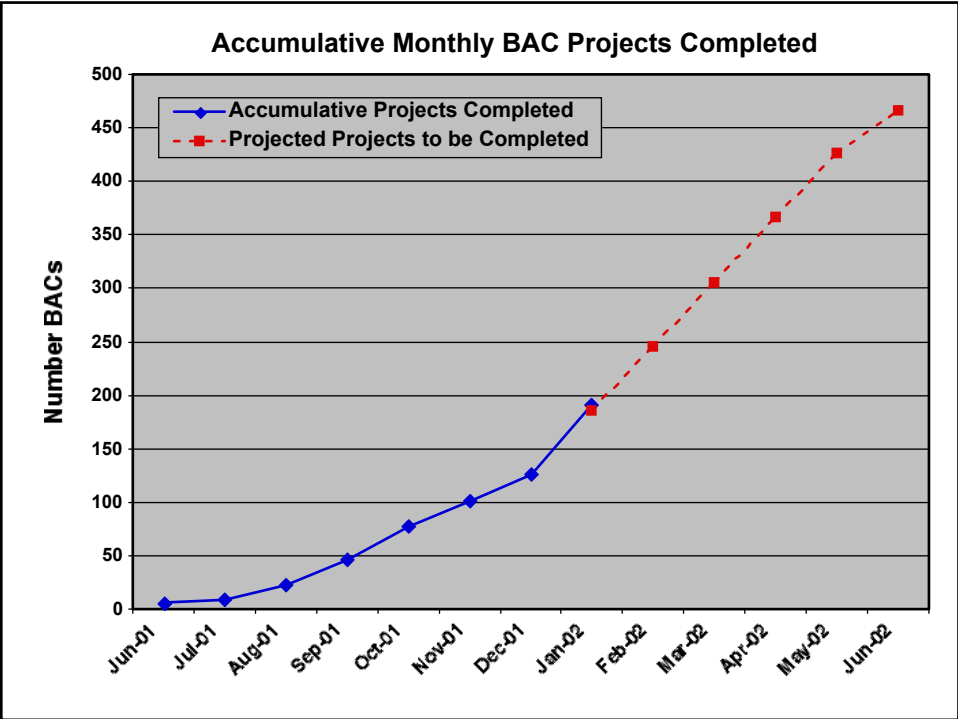
### Major Finishers on TPF (total of clone sizes)

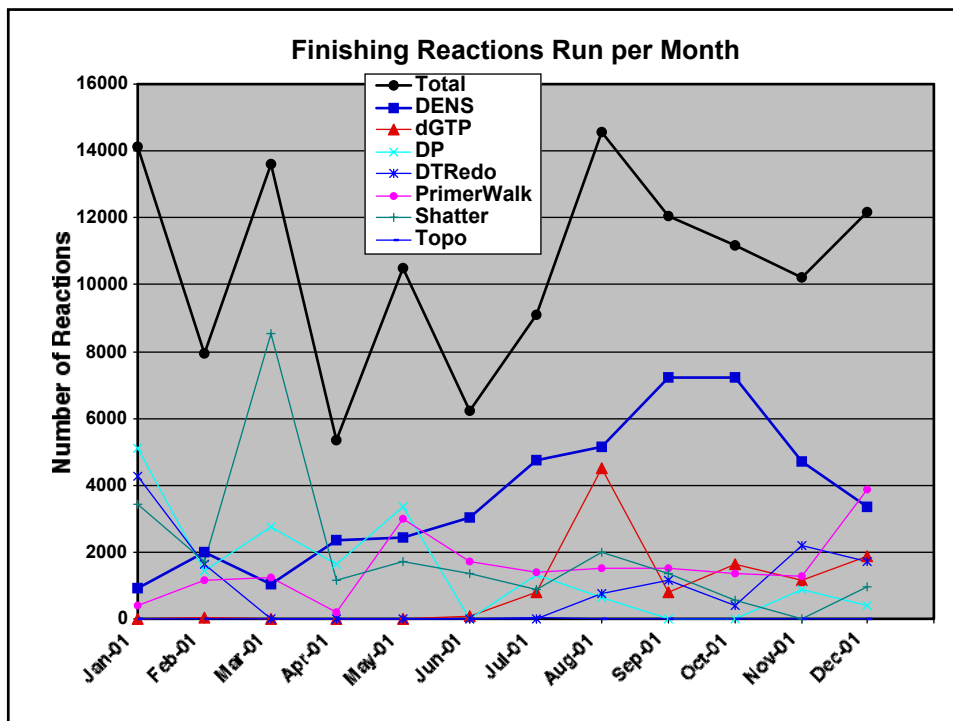
LANL	38,456,891
Stanford	13,157,867
TIGR	9,674,910
Sanger	1,442,639
Wash U	1,156,547




### Monthly Target Status









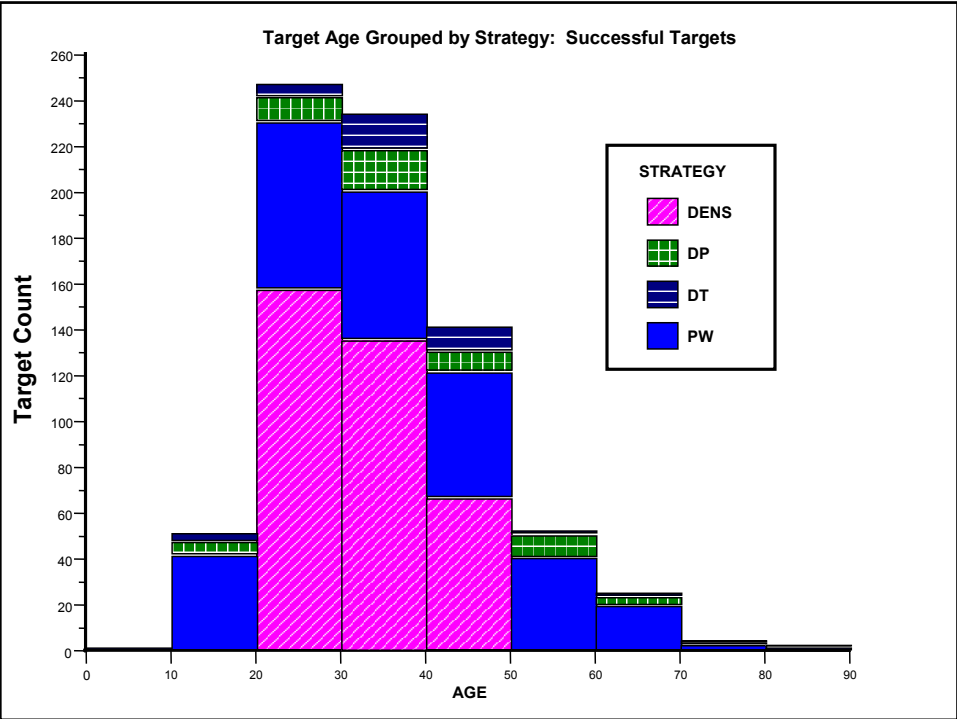
**JGI**  
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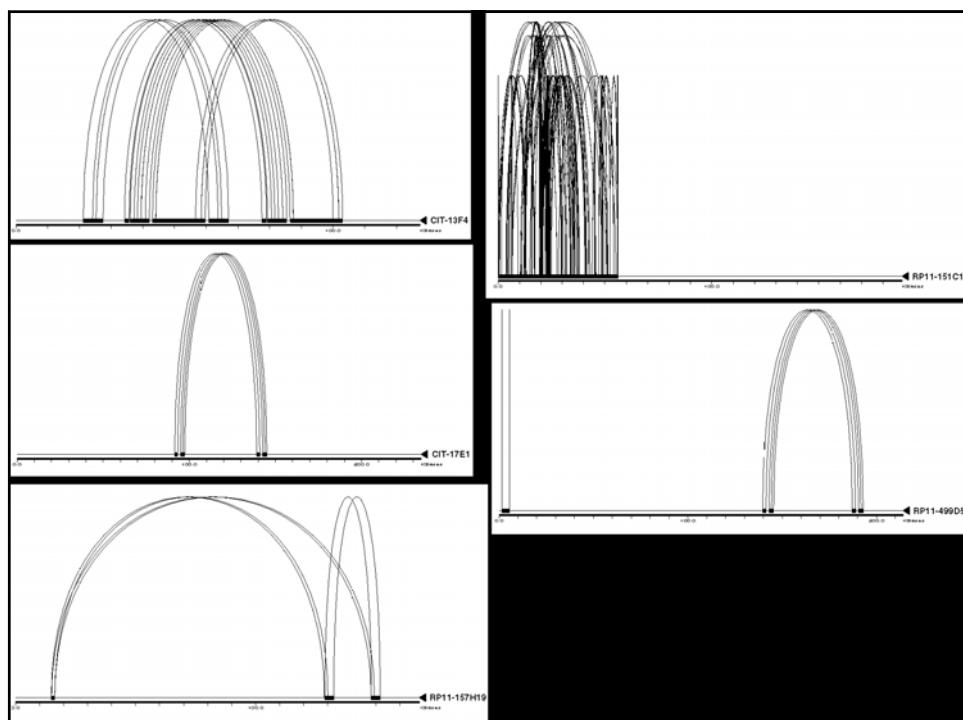
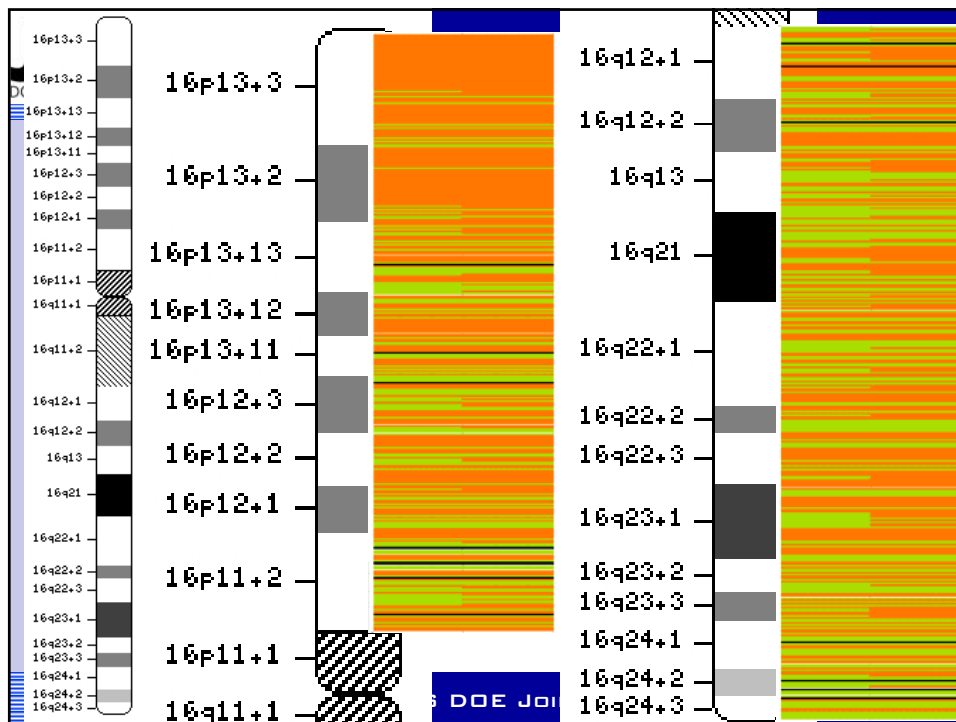
## Finishing Reads per Kb

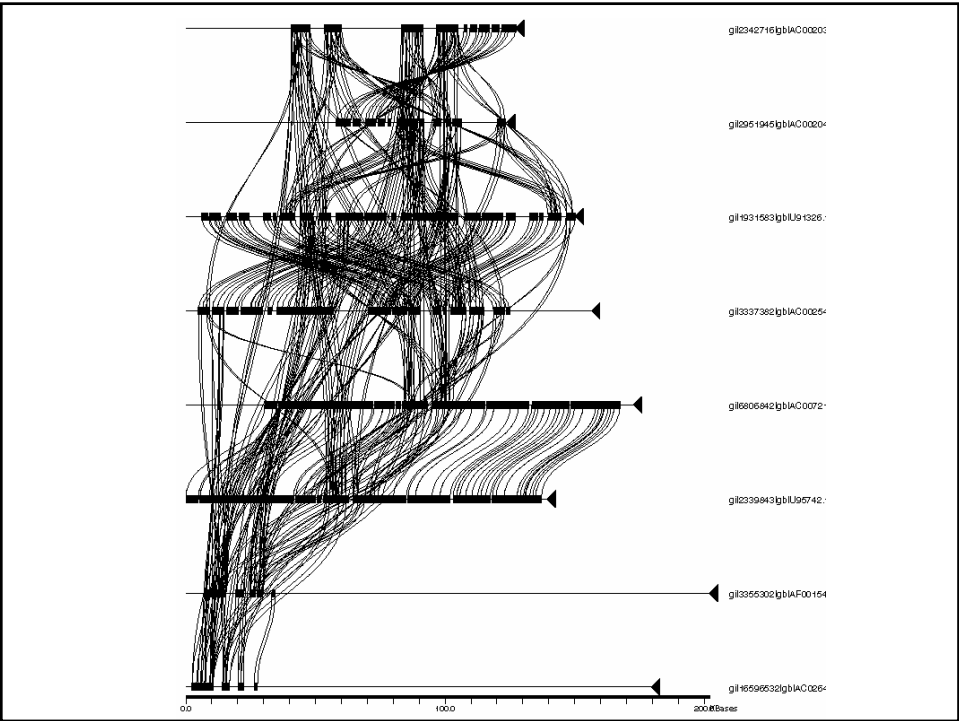
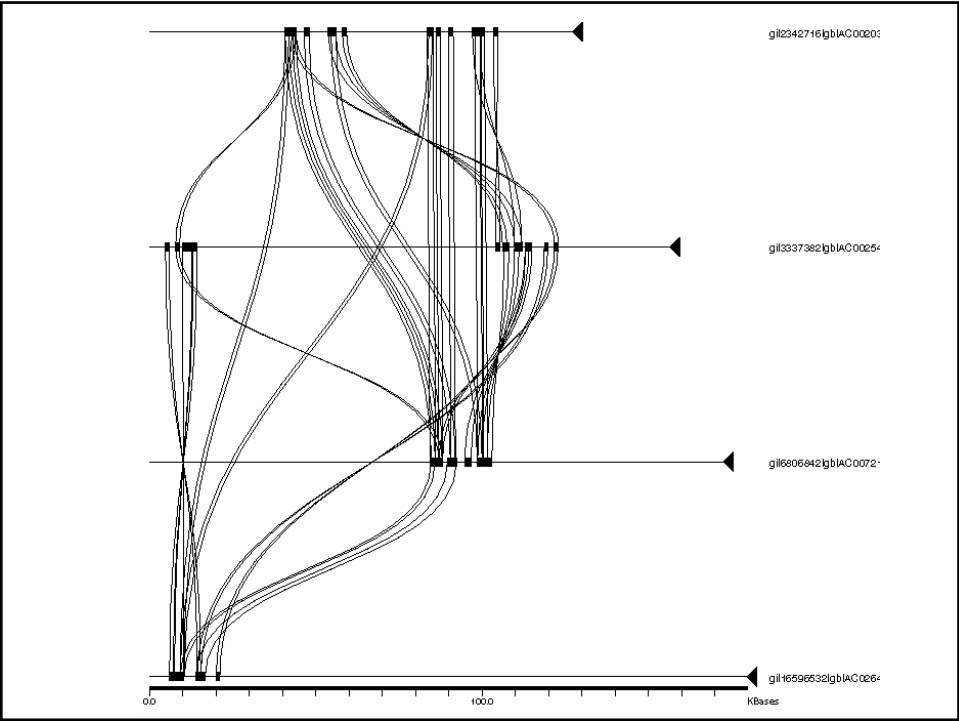
Draft/Finish	Min.	Avg.	Max.
LANL/LANL	0.3	3.4	7.2
JGI/LANL	0.1	3.2	12.0
G5/JGI/LANL	0.1	0.8	1.5

(Data from ~ 100 BAC projects finished in 2000-2001)

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- Larry Deaven

Marv Frazier

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